



Metabolic Flux Analysis in Mitochondria

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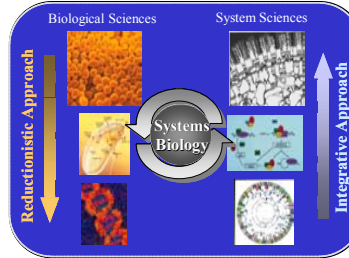


Abstract

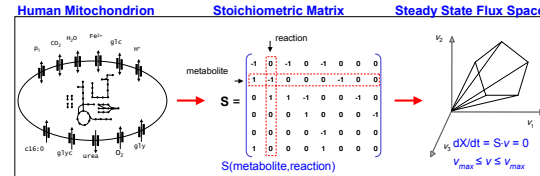
The emergence of high-throughput data has enabled the study of mitochondria as systems. We have reconstructed and characterized the human mitochondrial metabolic network based on proteomic and biochemical data. Linear programming and Monte Carlo sampling methods were applied to identify candidate steady states consistent with the imposed physiological and chemical observations. Analysis of equivalent optimal flux distributions, calculated with respect to each of the three metabolic functions, identified a group of flux distributions that are highly correlated, and thus are likely to be physiological relevant. Samples of steady-state flux distributions showed that the experimentally observed reduced activity of pyruvate dehydrogenase in diabetic and ischemic patients could be a result of stoichiometric constraints, and may not necessarily require enzymatic inhibition. Application of isotopomer data from isolated mouse hearts identified the fate of perfused [U-¹³C₆]glucose and [U-¹³C₃]pyruvate and flux redistribution at key substrate branch points.

Methods

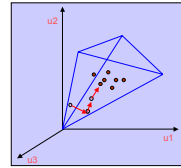
The constraint-based approach for analyzing reconstructed networks involves the application of a series of constraints arising from reaction stoichiometry, thermodynamics, enzymatic capacities, and regulatory and isotopomer balance constraints when they are available.



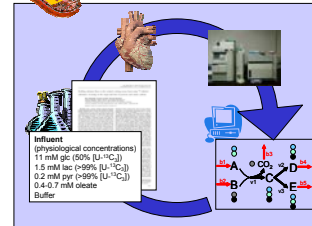
Analysis of equivalent optimal solutions



Monte Carlo sampling of solution space

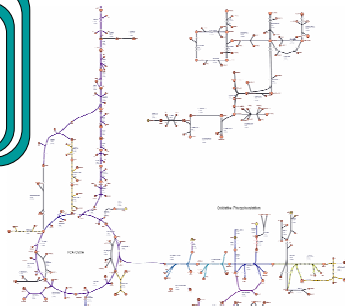
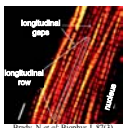
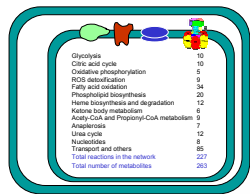
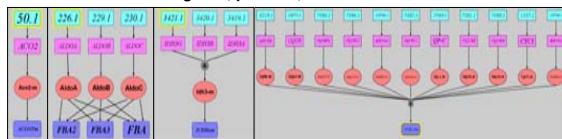


Application of isotopomer data



The mitochondrial network

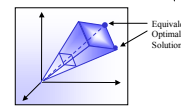
Association between genes, proteins, and reactions



Results

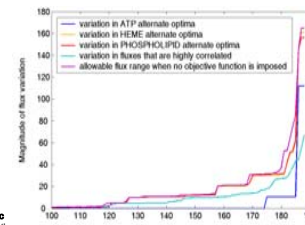
Metabolic objective functions

ATP production $Z = -1 \text{ ATP} - 1 \text{ H}_2\text{O} + 1 \text{ ADP} + 1 \text{ P}_i + 1 \text{ H}^+$
 Phospholipid biosynthesis $Z = 0.18 \text{ cardiolipin} + 0.34 \text{ phosphatidylethanolamine} + 0.43 \text{ phosphatidylcholine}$
 Heme biosynthesis $Z = \text{protoheme}$

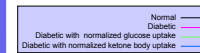


	ATP	Heme	Phospholipids
Total number of optima	7	6,208	23,403
Average reaction used per optimal solution	52.5	26.7	96.1
Reactions with variable fluxes	35	124	134
Reactions with constant fluxes	42	14	25
Reactions always used (shared constraint)	50	35	56
Reactions never used	127	65	54
Number of feasible optima per protein	9,360	33,600	161,972

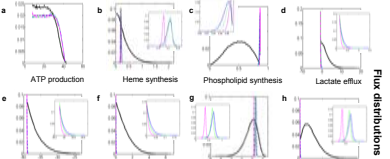
Reaction participation among equivalent optimal flux distributions calculated with respect to the three metabolic objective functions. Optima refer to the extreme points of the solution space that achieve the optimal value for the objective function, whereas feasible extreme points only satisfy the constraints of the linear programming problem.



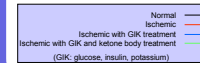
ATP production slightly reduced
 Excess fatty acid increases phospholipid synthesis
 Minimal pyruvate dehydrogenase activity
 Restricted overall network activity
 Effects of normalized glucose and ketone body uptake are positive but small.



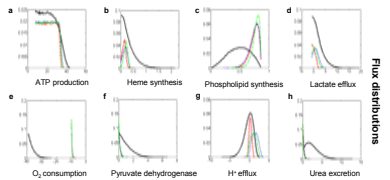
Diabetic condition



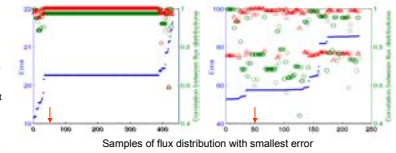
Moderate ischemia only slightly reduces ATP production
 Increased effluxes of lactate and H+
 Restricted overall network activity
 Treatments that directly activate pyruvate dehydrogenase will have minimal effect



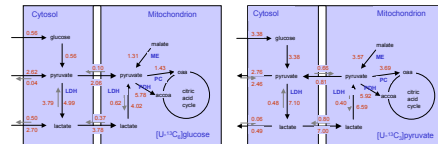
Ischemic condition



Error refers to the overall deviation between measured and predicted mass distribution.
 Correlation between the smallest error flux distribution and the remaining flux distributions.
 Correlation between the flux distribution marked at red arrow and the remaining.



Steady state reaction rates at pyruvate branch point



Conclusions

1. An *in silico* framework integrating multiple datasets is useful for studying mitochondrial metabolism under normal and stress conditions, and provides a basis for assessing effects of potential disease treatments.
2. Metabolic flux profiles with isotopomer data can uncover details about substrate utilization, substrate redistribution at network branch points, and quantitative information about enzyme activity.

References

1. Vo TD, Greenberg HJ, Palsson BO. 2004. Reconstruction and functional characterization of the human mitochondrial metabolic network based on proteomic and biochemical data. *J Biol Chem* 279(38):39532-40.
2. Thiele I, Price ND, Vo TD, Palsson B. 2004. Candidate metabolic network states in human mitochondria: impact of diabetes, ischemia, and diet. *J Biol Chem*. in press